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Synthesis and pharmacological evaluation of novel antinociceptive Nsubstituted-phenylimidazolyl-4-acylhydrazone derivatives

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Abstract

This paper describes recent results of design, synthesis and pharmacological evaluation of new N-heterocyclic functionalized N-acylhydrazone compounds (NAH), belonging to the N-substituted-phenylimidazolyl-4-acylhydrazone class (3a-o). These compounds were planned by applying the molecular hybridization strategy to propose the structural modifications on the previously described functionalized 2-methyl-imidazolyl-3-acylhydrazone class (2), which presented an important analgesic profile. This new series (3) was synthesized in order to investigate the possible pharmacophoric contribution of the N-heteroaromatic ring and N-acylhydrazone moieties to the analgesic activity. Compounds 3g and 3n are the most potent analgesic agents from this series, at the screening dose of 100 mg/kg *p.o.* and compounds 3e, 3j and 3o presented the best antiinflammatory properties at the same screening concentration.

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1. Introduction

In the course of an ongoing research program aiming at the design, synthesis and pharmacological evaluation of new bioactive N-acylhydrazone (NAH) derivatives [1-3], we described previously the analgesic and antiinflammatory profile of functionalized imidazo[1,2-a]pyridine 3-acylhydrazone derivatives (1) [1]. Next, we designed a second class of azaheterocyclic NAH derivatives, now belonging to 2-methyl-imidazolyl-3-acylhydrazone class (2) [4]. Series (2) was structurally planned by molecular simplification of 1 (Fig. 1) and presented the most important analgesic NAH derivatives obtained in the laboratory [5,6] when orally administered to mice in doses of 100 µM. These results were preliminary rationalized on the basis of the contribution of the less hydrophobic imidazole ring, compared with the previous series of NAH derivatives.

We wish describe in this paper a new class of imidazolyl NAH derivatives **3** following our studies on new azaheterocyclic *N*-acylhydrazone (*NAH*) derivatives as antinociceptive agents. These compounds were structurally planned by applying the molecular hybridization strategy on the previous class of 2-methylimidazolyl-3-acylarylhydrazone (**1**) derivatives and aminopyrine (**4**), a known pyrazolone with analgesic properties [7] (Fig. 1).

The rational basis of the molecular hybridization employed in the structural design of the new series **3** maintains the NAH moiety (sub-unit **A**, Fig. 1) and the imidazole ring presents in the most active compound of the previous series (**2**) [4]. In addition, we thought that it could be interesting to retain a similar substitution pattern around the imidazole nucleus and namely the NAH chain, with an *ortho*-monovalent isosteric substituent, i.e. C-5 methyl ($\sigma_{ortho} = 0.84$; MR = 5.65) in (**2**) and C-5 chloro ($\sigma_{ortho} = 0.76$; MR = 6.03) [8] in (**3**), such that the heterocyclic ring would retain a similar overall shape compared to the previously analgesic series **2**. Finally, we introduced the sub-unit **C** (Fig. 1) from

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Fig. 1. Design of N-substituted-phenylimidazolyl-4-acylhydrazone derivatives 3a-o.

pyrazolone (4), keeping the *N*-phenyl and *C*-dimethylamine substituent arriving to the structure of series 3. The selection of Ar group at end of imine double bond in this series was based on the previously described one as well as the nature of the **R** substituent at the *N*-phenyl ring [4–6].

2. Results and discussion

The synthesis of the new substituted NAH derivatives 3a-o is depicted in Scheme 1. The known N-substituted 5-chloro-2-(dimethylamino)imidazole-4-carboxaldehydes 5a-e, possessing different substituents at the Nphenyl moiety (R = H, Me, Cl and OMe), were prepared according to the procedure developed for construction of nitrogen heterocycles, under Vilsmeier conditions [9]. The oxidation of 5a-e with manganese dioxide and sodium cyanide in MeOH, afforded the desired methyl esters 6a - e [10], which were next converted to the corresponding hydrazides intermediates 7a-e by treatment with hydrazine hydrate (Table 1). Finally, compounds 7a-e were obtained by condensing with functionalized aromatic aldehydes, i.e. benzaldehyde, 4-dimethylaminobenzaldehyde, 4-bromobenzaldehyde, furan-2-carboxaldehyde and thiophene 2-carboxaldehyde, at reflux in methanol, to produce the desired Nsubstituted-phenylimidazolyl-4-acylhydrazones (3a-o)in good yields, as illustrated in Table 2.



Scheme 1. (a) NaCN, MnO₂, MeOH, rt, 12h, 84–92%. (b) NH₂NH₂. H₂O 80%, EtOH, reflux, 8–10h, 70–93%. (c) Ar-CHO, EtOH, HCl (cat.), reflux, 2h, 85–96%.

The structure of these stable and crystalline compounds were fully characterized by the usual methods. (IR, ¹H, ¹³C NMR). IR spectra of these new NAH derivatives presented the N–H absorption at 3304–3489 cm⁻¹, the C=O at 1655–1748 cm⁻¹ and C=N at 1570– 1645 cm⁻¹ stretching bands (Table 2). The *E*-configuration of the C=N double bond in the compounds **3a–o**

Table 1 Yields, IR and ¹H NMR spectral data of compounds 6a-d and 7a-d



Compound	R	Yield (%)	M.p. (°C)	$IR^{b}(cm^{-1})$ C=O	IR (cm ⁻¹) (N-H)	¹ H NMR and δ (ppm)
6a	Н	92	75-76 ^a	1709	-	7.58–7.50 (m, 3H, H3 and H4'), 7.36–7.30 (m, 2H, H2'), (s, 3H, OCH ₂) 2.64 (s, 6H, N(CH ₂) ₂)
6b	CH ₃	90	$135{-}136$ ^a	1709	-	7.21 (d, 2H, $J = 8.3$ Hz, H3'), 7.12 (d, 2H, $J = 8.3$ Hz, H2'), 3.83 (s, 3H, OCH ₃), 2.57 (s, 6H, N(CH ₃) ₂), 2.36 (s, 3H, CH ₃)
60	OCH ₃	84	$145 - 150^{a}$	1677	—	7.50 (d, 2H, <i>J</i> = 8.8 Hz, H2'), 7.36–7.26 (m, 2H, H3'), 3.91 (s, 3H OCH ₃), 3.91 (s, 3H, O(CH ₃ aromatic), 2.64 (s, 6H, N(CH ₃) ₂)
6d	Cl	85	188–189 ^a	1720	_	7.64 (d, 2H, <i>J</i> = 8.8 Hz, H3'), 7.47 (d, 2H, <i>J</i> = 8.8 Hz, H2'), 8.70 (br, 1H, NHNH ₂), 4.33 (br, 2H, NHNH ₂), 2.55 (s, 6H, N(CH ₃) ₂)
7a	Н	74	195-196	1653	3301; 3206	7.56–7.43 (m, 5H, aromatic), 3.35 (br, 3H, NHNH ₂), 2.53 (s, 6H, N(CH ₃) ₂)
7b	CH3	93	204-205	1653	3301; 3206	7.34 (d, 2H, <i>J</i> = 7.3 Hz, H3'), 7.28 (d, 2H, <i>J</i> = 7.3 Hz, H2'), 8.78 (br, 1H, NHNH ₂), 4.50 (br, 2H, NHNH ₂), 2.50 (s, 6H, N(CH ₃) ₂), 2.39 (s, 3H, CH ₃)
7c	OCH ₃	70	194–196	1654	3430; 3299	7.32 (d, 2H, $J = 9.0$ Hz, H2'), 7.10 (d, 2H, $J = 9.0$ Hz, H3'), 8.32 (br, 1H, NHNH ₂), 4.37 (br, 2H, NHNH ₂), 3.80 (s, 3H, OCH ₃), 2.53 (s, 6H, N(CH ₃) ₂)
7d	Cl	77	224-226	1656	3297; 3204	7.64 (d, 2H, $J = 8.8$ Hz, H3'), 7.47 (d, 2H, $J = 8.8$ Hz, H2'), 8.70 (br, 1H, NHNH ₂), 4.33 (br, 2H, NHNH ₂), 2.55 (s, 6H, N(CH ₃) ₂)

^a Purified by silica gel colunm chromatography.
 ^b Obtained from KBr plates.

Table 2

N-Substituted-phenylimidazolyl-4-acylarylhydrazone derivatives 3a-o



Compound	R	W/X	Yield (%)	M.p. (°C)	Molecular formula ^a	Molecular weight	$R (cm^{-1})$
3a	Н	Н	91	185	C ₁₉ H ₁₈ ClN ₅ O	367.84	3303 (N-H), 1655 (C=O), 1574 (C=N)
3b	Н	$N(CH_3)_2$	95	176-179	C21H23ClN6O	410.90	3344 (N-H), 1661 (C=O), 1600 (C=N)
3c	Н	Br	91	171 - 174	C19H17BrClN5O	446.73	3304 (N-H), 1674 (C=O), 1590 (C=N)
3d	CH_3	Н	85	184-185	C20H20ClN5O	381.86	3314 (N-H), 1672 (C=O), 1571 (C=N)
3e	CH_3	$N(CH_3)_2$	96	190-191	C22H25ClN6O	424.93	3304 (N-H), 1684 (C=O), 1599 (C=N)
3f	CH_3	Br	89	169 - 170	C ₂₀ H ₁₉ BrClN ₅ O	460.76	3281 (N-H), 1673 (C=O), 1590 (C=N)
3g	CH_3	0	90	200-201	C18H18ClN5O2	371.82	3442 (N-H), 1669 (C=O), 1572 (C=N)
3h	CH_3	S	84	250-252	C ₁₈ H ₁₈ ClN ₅ OS	387.89	3424 (N-H), 1672 (C=O), 1573 (C=N)
3i	OCH ₃	Н	90	184-185	C ₁₉ H ₁₈ ClN ₅ O	402.28	3210 (N-H), 1678 (C=O), 1556 (C=N)
3j	OCH ₃	$N(CH_3)_2$	95	199-200	C ₂₁ H ₂₂ Cl ₂ N ₆ O	445.35	3218 (N-H), 1747 (C=O), 1557 (C=N)
3i	OCH ₃	Br	70	183-185	C19H16BrCl2N5O	481.18	3303 (N-H), 1748 (C=O), 1589 (C=N)
3m	Cl	Н	63	193-194	$C_{20}H_{20}ClN_5O_2$	397.86	3231 (N-H), 1655 (C=O), 1570 (C=N)
3n	Cl	$N(CH_3)_2$	90	177 - 178	C ₂₂ H ₂₅ ClN ₆ O ₂	440.93	3271 (N-H), 1746 (C=O), 1645 (C=N)
30	Cl	Br	95	175 - 176	$C_{20}H_{19}BrClN_5O_2$	476.76	3297 (N-H), 1747 (C=O), 1661 (C=N)

 $^a\,$ The analytical results for C, H, N were within $\pm 0.4\%$ of calculated values.

Table 3 ¹H NMR spectral data at 200 MHz (DMSO-d₆) for N-substituted-phenylimidazolyl-4-acylarylhydrazone derivatives 3a-o



Compound δ (ppm)

	H-2′	H-3′	H-4′	N=CH	H-2''	H-3"	H-4''	N-H	Other
3a	7.53-7.44 (m)	7.53-7.44 (m)	7.53–7.44 (m)	8.57 (s)	7.62-7.56 (m)	7.73–7.69 (m)	7.73-7.69 (m)	11.16 (s)	2.60 (s) N(CH ₃) ₂ (ring)
3b	7.58-7.48 (m)	7.53-7.48 (m)	7.53–7.48 (m)	8.38 (s)	7.53 (d) <i>J</i> = 8.3 Hz	6.76 (d) $J = 8.3$ Hz	_	10.84 (s)	2.59 (s) N(CH ₃) ₂ (ring), 2.96 (s) N(CH ₃) ₂
3c	7.62-7.46 (m)	7.62-7.46 (m)	7.62–7.46 (m)	8.55 (s)	7.65 (s)	7.65 (s)	-	11.30 (s)	2.61 (s) N(CH ₃) ₂
3d	7.30 (d) <i>J</i> = 8.9 Hz	7.39 (d) <i>J</i> = 8.9 Hz	_	8.55 (s)	7.73-7.68 (m)	7.48-7.39 (m)	7.48-7.39 (m)	11.00 (s)	2.40 (s) CH ₃ , 2.63 (s) N(CH ₃) ₂
3e	7.30 (d) $J = 8.4$ Hz	7.39 (d) <i>J</i> = 8.4 Hz	_	8.35 (s)	7.52 (d) $J = 9.0$ Hz	6.75 (d) $J = 9.0$ Hz	-	10.66 (s)	2.40 (s) CH ₃ , 2.60 (s) N(CH ₃) ₂ (ring), 2.97 (s) N(CH ₃) ₂
3f	7.31 (d) $J = 8.5$ Hz	7.39 (d) <i>J</i> = 8.5 Hz	_	8.52 (s)	7.64 (s)	7.64 (s)	-	11.06 (s)	2.40 (s) CH ₃ , 2.60 (s) N(CH ₃) ₂
3g	7.30 (d) $J = 8.2$ Hz	7.39 (d) <i>J</i> = 8.2 Hz	_	8.46 (s)	6.85 (d) $J = 3.2$ Hz	6.65 (d) $J = 3.2$ and 1.2 Hz	7.82 (d) $J = 1.2$ Hz	11.00 (s)	2.39 (s) CH ₃ , 2.59 (s) N(CH ₃) ₂
3h	7.27 (d) $J = 8.2$ Hz	7.38 (d) $J = 8.2$ Hz	_	8.62 (s)	7.52 (d) $J = 5.1$ Hz	7.08 (dd) $J = 5.1$ and 3.6 Hz	7.26 (d) $J = 3.6$ Hz	11.05 (s)	2.38 (s) CH ₃ , 2.56 (s) N(CH ₃) ₂
3i	7.36 (d) $J = 8.9$ Hz	7.11 (d) <i>J</i> = 8.9 Hz	_	8.46 (s)	7.72-7.68 (m)	7.45-7.40 (m)	7.45-7.40 (m)	11.20 (s)	2.60 (s) N(CH ₃) ₂ , 3.82 (s) OCH ₃
3j	7.35 (d) $J = 8.9$ Hz	7.10 (d) <i>J</i> = 8.9 Hz	_	8.35 (s)	7.51 (d) $J = 8.9$ Hz	6.74 (d) $J = 8.9$ Hz	_	10.71 (s)	2.61 (s) N(CH ₃) ₂ (ring), 2.96 (s) N(CH ₃) ₂
3i	7.36 (d) $J = 9.0$ Hz	7.11 (d) $J = 9.0$ Hz	_	8.52 (s)	7.64 (s)	7.64 (s)	_	11.11 (s)	2.62 (s) N(CH ₃) ₂ , 3.83 (s) OCH ₃
3m	7.54 (d) $J = 8.5$	7.68 (d) $J = 8.5$	_	8.60 (s)	7.70-7.66 (m)	7.56-7.44 (m)	7.56-7.44 (m)	11.20	2.60 (s) N(CH ₃) ₂
3n	7.54 (d) $J = 8.5$	7.68 (d) $J = 8.5$	_	8.34 (s)	7.52 (d) $J = 8.9$ Hz	6.76 (d) $J = 8.9$ Hz	_	10.84	2.60 (s) N(CH ₃) ₂ , 2.97 (s) N(CH ₃) ₂
30	7.54 (d) $J = 8.9$ Hz	7.69 (d) $J = 8.9$ Hz	_	8.35 (s)	7.65 (s)	7.65 (s)	_	(s) 11.30 (s)	2.61 (s) N(CH ₃) ₂

Table 4 13 C NMR spectral data at 50 MHz (DMSO-d₆) for *N*-substituted-phenylimidazolyl-4-acylarylhydrazone derivatives **3a**-**o**





Fig. 2. (E)-configuration of derivatives 3a-o.

was determined by carefully ¹H, ¹³C NMR analysis (Tables 3 and 4) [11,12]. In fact, the (*E*)-diastereomers of NAH presented the N=CH signal at 7.83–8.33 ppm. By applying the NOE differential experiments we were able to detect the steric vicinity of the N–H and N=CH hydrogens. For instance, the irradition of the N–H signal (10.66 ppm) expressively enhanced (22%) the N=CH hydrogen signal, occurring at 8.35 ppm (Fig. 2).

The analgesic and anti-inflammatory activities of the new N-substituted-phenylimidazolyl-4-acylhydrazone derivatives (3a-o) were evaluated using the acetic acid-induced mice abdominal constrictions test [13] and the carrageenan-induced rat paw edema test [14], respectively. The results are disclosed in Tables 5 and 6,

respectively. All compounds were studied with a concentration of $100 \ \mu mol/kg$.

The most potent analysic compound $3g (R = CH_3)$ and R = 2-furyl) (Table 5) showed 45.8% of inhibition of the induced contortions, higher than dipyrone used as standard (35.9%) at the same dose, followed by compound **3n** ($\mathbf{R} = \mathbf{Cl}$ and $\mathbf{W} = \mathbf{N}(\mathbf{CH}_3)_2$), which presented 35.2% of inhibition. Interestingly, the corresponding 5metylimidazolyl-4-acylhydrazone derivative (2), described in the previous series [4] presented 72% of inhibition, suggesting that the presence of 2-N-dimethylamine unit and the N-phenyl at the imidazole ring was detrimental to the analgesic activity of this series. In addition, the nor-chloro congener (3b) of 3n, presented only 18.3% of inhibition, and the corresponding paramethyl isostere (3e) was practically inactive. These results seem to indicate that the analgesic activity identified for derivatives possessing the N-dimethylphenyl unit at the imine terminus (i.e. 3n, 3b, 3j, 3e) is related to a combination of π_{arom} and σ parameters of the para-substituent of the N-phenyl ring. In contrast, the analgesic activity of other imidazole derivatives (3a, 3c-d, 3f-i, 3l-m, 3o) seems to be related to a balance of different structural contributions.

Table 5

Effect of N-substituted-phenylimidazolyl-4-acylarylhydrazone derivatives 3a-o and dipyrone on the inhibition of abdominal constrictions induced by acetic acid (0.6%, *i.p.*) in mice



Compound ^a	R	W/X	N ^b	Constrictions count	Inhibition (%) ^c	Relative activity ^d
Vehicle control (arabic gum 5%)	_	_	10	79.8 ± 4.3	_	_
Dipyrone	_	-	10	38.7 ± 4.9	35.9	1.00
3a	Н	Н	11	56.6 ± 5.5	28.3	0.79
3b	Н	$N(CH_3)_2$	12	64.5 ± 4.9	18.3	0.51
3c	Н	Br	10	72.0 ± 4.3	8.9 ^e	0.25
3d	CH_3	Н	8	66.6 ± 3.7	15.7 ^e	0.44
3e	CH_3	$N(CH_3)_2$	9	72.8 ± 3.4	8.4 ^e	0.23
3f	CH_3	Br	7	71.0 ± 4.2	10.1 ^e	0.28
3g	CH_3	0	9	42.8 ± 5.0	45.8 ^f	1.28
3h	CH ₃	S	10	75.8 ± 6.2	1.3 ^e	0.04
3i	OCH_3	Н	8	69.5 ± 3.9	12.1 ^e	0.34
3j	OCH_3	$N(CH_3)_2$	10	64.5 ± 5.5	18.3 ^e	0.51
31	OCH_3	Br	9	58.3 ± 6.7	26.2 ^f	0.73
3m	Cl	Н	10	87.2 ± 4.1	-10.3^{e}	-0.29
3n	Cl	$N(CH_3)_2$	13	51.2 ± 4.9	35.2 ^f	0.98
30	Cl	Br	14	61.43 ± 5.9	22.2 ^f	0.62

^a All test compounds were administered at a dose of 100 µmol/kg p.o.

^b N = number of animals.

e Not significant.

 $^{\rm f}$ p < 0.05 (Student's t-test). Results are expressed as mean \pm SEM.

^c % inhibition obtained by comparison with vehicle control group.

^d Analgesic activity relative to dipyrone.

Table 6

Effect of N-substituted-phenylimidazolyl-4-acylarylhydrazone derivatives 3a-o and indomethacin in the inhibition of carrageenan-induced rat paw edema



Compound ^a	R	W/X	N ^b	Volume variation (μ L) ^c	Inhibition (%) ^d
Vehicle control (arabic gum 5%)	_	_	10	478.0 ± 30.0	_
Indomethacin	_	-	9	228.1 ± 31.9	52.3
3a	Н	Н	10	341.3 ± 43.5	28.6 ^f
3b	Н	$N(CH_3)_2$	11	435.1 ± 36.7	90.0 ^e
3c	Н	Br	10	452.5 ± 25.5	5.3 °
3d	CH_3	Н	8	404.3 ± 38.5	15.5 °
3e	CH_3	$N(CH_3)_2$	6	299.3 ± 26.2	37.5 ^f
3f	CH_3	Br	7	431.7 ± 35.0	9.7 °
3g	CH_3	0	6	343.5 ± 41.2	28.2 ^f
3h	CH_3	S	6	399.2 ± 56.2	16.5 °
3i	OCH_3	Н	6	390.2 ± 31.7	18.4 °
3j	OCH ₃	$N(CH_3)_2$	6	334.0 ± 40.0	30.1 ^f
31	OCH_3	Br	6	341.2 ± 44.9	28.6 ^f
3m	Cl	Н	7	341.7 ± 47.4	28.6 ^f
3n	Cl	$N(CH_3)_2$	7	345.6 ± 36.7	27.7 ^f
30	Cl	Br	6	325.3 ± 49.4	32.0 ^f

^a All test compounds were administered at a dose of 100 µmol/kg p.o.

^b N = number of animals.

^c Volume variation is the difference between the volumes of carrageenan- and saline-treated paws at the 3 h after carrageenan injection paw.

 $^{\rm d}\,$ Percentage of inhibition obtained by comparison with the vehicle control group.

^e Not significant.

 $^{\rm f}$ p < 0.05 (Student's t-test). Results are expressed as mean \pm SEM.

The results of the anti-inflammatory bioassay for series 3, also measured with a concentration of 100 umol/kg (Table 6) indicated that derivatives possessing the para-dimethylaminophenyl unit at the imine end were, with exception of 3b, was the most active ones, inhibiting the edema formation for 3 h after the induction in 37.5%. For instance, the N-para-methoxyphenyl analogue 3j presented 30.1% of edema inhibition followed by the corresponding N-parachlorophenyl isoster (3n) that presenting 27.7% of inhibition. In addition, the bis-halogenated derivative 30 presented 32.0% of edema inhibition. The data disclosed in Table 6 indicate that all compounds having the N-para-chlorophenyl moiety were active in this essay. For instance, compounds 3m and 3n presented an edema inhibition of 28% The anti-inflammatory profile of the most analgesic derivative of this series, compound 3g was measured as 28.2% and was similar to the previously described compound of series 2, which presented 33% of edema inhibition at the same concentration [4].

Furthermore, the effect of the most anti-inflammatory derivatives (3a, 3e, 3g, 3l-m and 3o) in the gastric

mucosa was also investigated, indicating a very poor ulcerative behavior when essayed at the concentration of $300 \ \mu mol/kg$ (data not shown).

3. Conclusions

These results identified the 2-furyl compound (3g) as the most attractive compound, with the best antinociceptive profile, and present an attracting anti-inflammatory behavior for this new series of imidazole NAH derivatives (3).

4. Experimental

4.1. Chemistry

M.p.s were determined with a Thomas Hoover apparatus and are uncorrected. ¹H NMR spectra, unless otherwise stated, were determined in deuterated Me₂SO containing ca. 1% Me₄Si as an internal standard using a Bruker AC 200 spectrometer at 200 MHz. Splitting patterns are as follows: s, singlet; d, doublet; dd, double doublet; br, broad; m, multiplet. ¹³C NMR was determined using the same spectrometer described above at 50 MHz, using deuterated Me₂SO as internal standard. IR spectra were obtained using a Bruker IFS66 spectrophotometer by using KBr plates. Microanalysis data was obtained using a Perkin–Elmer 240 analyzer, using a Perkin–Elmer AD-4 balance.

The progress of all reactions was monitored by TLC performed on $2.0 \times 6.0 \text{ cm}^2$ aluminum sheets precoated with silica gel 60 (HF-254, E. Merck) to a thickness of 0.25 mm. The developed chromatograms were viewed under ultraviolet light at 254 nm. For column chromatography E. Merck silica gel (60–200 mesh) was used. The N-substituted 5-chloro-2-(dimethylamino)imidazole-4-carboxaldehydes **5a**–e were prepared by known procedure in the literature [9].

4.1.1. General procedure for the preparation of methyl N-substituted 5-chloro-2-(dimethylamino)imidazole-4carboxylate (6a-e)

To a mixture of 1.45 mmol of N-substituted 5-chloro-2-(dimethylamino)imidazole-4-carboxaldehydes 5a-e in methanol (18 ml), and 0.35 g (7.14 mmol) of NaCN, activated MnO₂ 1.60g (18.4 mmol) was added. The reaction was stirred at room temperature for 12 h and then the suspension was filtered through Celite[®] and treated with CH₂Cl₂ (3 × 20 ml). The organic layers were joined, dried over anhydrous Na₂SO₄, filtered and evaporated under reduced pressure to furnish crude methyl esters 6a-e, which were purified by flash column chromatography using 80% *n*-C₆H₁₄-EtOAc as eluent (see Table 1).

4.1.2. General procedure for preparation of the N-substituted-phenylimidazolyl-4-carbohydrazide derivatives (7a-e)

A solution the appropriate methyl ester derivates 6a-e (3.41 mmol) and 3.2 ml of 80% hydrazine monohydrate in 16 ml of EtOH, was stirred at reflux for 8–10 h. After this time, the reaction mixture was concentrated under reduced pressure and the colored solid formed was collected by filtration, washed with cold water and dried under vacuum to give the desired imidazolyl hydrazides 7a-e, as showed in Table 1.

4.1.3. General procedure for the preparation of the N-substituted-phenylimidazolyl-4-acylhydrazone derivatives (3a-o)

To a solution of hydrazide derivatives 7a-e (0.50 mmol) in 15 ml of EtOH, was added an equimolar amount of the appropriates aromatic aldehydes in the presence catalytic amount of 37% aqueous HCl. The reaction was stirred for 2 h, at reflux. Then, solvent was evaporated and the colored precipitate was collected by filtration, washed with cold water and dried under

vacuum. The imidazolylhydrazone derivates 3a-o were purified in column flash chromatography using 50% *n*-C₆H₁₄-EtOAc as eluent (Tables 3 and 4).

4.2. Pharmacology

4.2.1. Analgesic activity

The analgesic activity was determined in vivo by using abdominal constriction test induced by acetic acid 0.6%(0.1 ml/10g) in mice [11]. Albino mice of both sexes (18-23 g) were used. Compounds were administered orally (100 µmol/kg; 0.1 ml/20 g) as a suspension in 5% arabic gum in saline (vehicle). Dipyrone (100 µmol/kg) was used as the standard drug under same conditions. Acetic acid solution was administered *i.p.* One hour after administration of the NAH compounds 3a-o. Ten minutes after *i.p.* injection of the acetic acid solution, the number of constrictions per animal was recorded for 20 min. Control animals received on equal volume of vehicle. Analgesic activity was expressed as percentage of inhibition of constrictions when compared with the vehicle control group. Results are expressed as the mean \pm SEM of *n* animals per group. The data were statistically analyzed by Student's *t*-test for significance level of * $\rho < 0.05$.

4.2.2. Antiinflammatory activity

The antiinflammatory activity was determined in vivo using the carrageenan induced rat paw edema test according to Pereira and coworkers [12]. Albino rats of both sexes (150-200 g) were used. Compounds were administered orally 100 µmol/kg (0.1 ml/20g) as a suspension in 5% arabic gum in saline (vehicle). Control animals received on equal volume of the vehicle. One hour after, the animals were then injected with either 0.1 ml of 1% carrageenan solution in saline (0.1 mg/paw) and sterile saline (NaCl 0.9%), into the subplantar surface of one of the hind paws, respectively. The paw volumes were measured using a glass plethysmometer coupled to a peristaltic pump, 3 h after the subplantar injection. The edema was calculated as the volume variation between the carrageenan and saline-treated paw. Indomethacin (100 µmol/kg) and was used as standard in the same conditions. Antiinflammatory activity was expressed as percentage of inhibition of the edema when compared with vehicle control group.

Results are expressed as the mean \pm SEM of *n* animals per group. The data were statistically analyzed by Student's *t*-test for significance level of * $\rho < 0.05$.

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